

Effect of Electrolysis Treatment Time, Field Strength and Recycling Flowrate on *Ankistrodesmus* sp. Lipid Extraction

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The study discusses the effect of Electrolysis Ereatment (ET) parameters focussed on the Electrolysis Treatment Time (ETT), Electrolysis Field Strength (EFS) and Electrolysis Recycling Flowrate (ERF) towards *Ankistrodesmus* sp. lipid extraction prior to the solvent extraction method. In the ET system selection experiment, the highest lipid content (13.05 ± 0.57 % wt.) attained was from ET system 3 (21V/cm, 140ml/min and 25 min). The three ET parameters were then optimized. The highest lipid content and methyl palmitate gained in ETT was 15.83 ± 0.68 % wt. and $0.440 \pm 0.046\mu\text{g/ml}$ at 20 min, respectively. Meanwhile, the highest lipid content and methyl palmitate obtained in EFS was 15.46 ± 0.78 % wt. and $0.629 \pm 0.058\mu\text{g/ml}$ at 21V/cm, respectively. As for ERF, the highest lipid content and methyl palmitate attained was 15.60 ± 0.67 % wt. and $0.390 \pm 0.032\mu\text{g/ml}$ at 180ml/min, respectively. In comparison to *Ankistrodesmus* sp. Without Electrolysis Treatment (WOET), the lipid extraction of *Ankistrodesmus* sp. treated With Electrolysis Treatment (WET) can be improved and had resulted in the increasing of extraction with 1.34, 1.38 and 1.40 times higher in ETT, EFS and ERF, respectively.

Keywords: *Ankistrodesmus* sp.; electrolysis treatment; lipid extraction; lipid content; methyl palmitate

I. INTRODUCTION

It is undeniable that humans are still relying on fossil fuel resources for transportation, heat generating and electricity purpose. The concerning part is that all of these natural resources soon will be diminished and will not be adequate to support the people's necessities worldwide. Moreover, it takes millions of years to wait for these fossil fuels to form again. For that reason, seeking other alternative energy for fossil fuel replacement is crucial and urgent. Biodiesel is a renewable and sustainable energy that can lessen fossil fuels utilisation as well as reduces the global warming impact, which is primarily contributed

from fossil fuels combustion (IEA, 2017). To date, the interest in using microalgae-based biodiesel has become more significant. High lipid content, rapid growth rate, less land for cultivation, flexible to grow under various climates, and none food security related are some of the unique features of microalgae (Joannes *et al.*, 2016; Mansa *et al.*, 2018a, 2012). In this study, *Ankistrodesmus* sp. was used because it has shown a very promising feedstock for biodiesel. This green freshwater microalga is capable of producing lipid around 32.4 to 57.7 % wt as well as enrich with monosaturated fatty acids such as palmitic acid (Sukkrom *et al.*, 2014).

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The microalga's lipid is one of the secret ingredients in biodiesel formation. The lipid is stored inside the microalga and is sturdily protected by its rigid cell wall, which is composed of cellulose, lipid, glycoprotein, polysaccharide and protein (Lee *et al.*, 2017). In other words, the cell wall is the major bottleneck in microalga lipid extraction. Hence, the cell wall must be broken to facilitate the lipid extraction process (acylglycerols and free fatty acids). Therefore, lipid extraction is relatively a challenging stage. Some researchers had used mechanical cell wall disruptions (e.g. high-pressure homogenizer, hydrodynamic, ultrasonication, microwave, Pulse Electric Field (PEF) and hydrothermal liquefaction) or non-mechanical cell wall disruptions (e.g. acid treatment, ionic liquid, osmotic shock, solvent extraction, supercritical fluid extraction, oxidation, surfactant, enzymatic lysis and algicidal treatment) to extract the lipid from microalgae (Günerken *et al.*, 2015; Joannes *et al.*, 2015b). Nonetheless, some drawbacks such as energy intensive, upscaling, time-consuming, chemicals safety, high capital investment, costly raw materials and highly precarious involving electrical based extraction (Mubarak *et al.*, 2015). Thus, numbers of researchers have combined mechanical and non-mechanical methods to improve lipid extraction efficiency.

Apart from these extraction methods, another recent approach which is using an Electrolysis Treatment (ET) as a prelude for microalgae lipid extraction (Mansa *et al.*, 2018a). The benefits of ET such as it uses lower voltage range (~ 6-30V), simple operation, cheaper, lighter, easy to construct and clean. Conventionally, ET method is used in microalgae harvesting with an electrolyte addition and typically conducted under a batch system (Baierle *et al.*, 2015; Uduman *et al.*, 2011; Zhou *et al.*, 2016). The process of collecting the microalga cells using ET is called electrophoresis. In nature, microalga cells are negatively charged, and the cells will move to the anode (positive terminal) and then coagulate due to the potential difference between the anode and cathode (Uduman *et al.*, 2010). In other theory, exposing the microalga cells to electric fields at a certain level (higher than the cell membrane critical strength) will cause the cell to experience irreversible or reversible cell electroporation. Irreversible cell electroporation is the condition where the cell membrane is unable to reseal back to its original form whereas, reversible cell electroporation is the opposite condition of irreversible cell electroporation (Joannes *et*

al., 2015a, 2015b; Mansa *et al.*, 2018b). When the cell membrane is broken, the valuable components like lipid, protein and carbohydrate will be exposed to the surrounding and easily can be extracted. In the meantime, only a few available literatures were published with regards to microalga lipid extraction using the ET method (Daghrir *et al.*, 2014; Guldhe *et al.*, 2016; Hua *et al.*, 2016; Misra *et al.*, 2015, 2014). This method is still under development and is considered to be a new approach to microalga lipid extraction.

Henceforth, as the continuity from the preliminary study conducted by Mansa *et al.* (2018b), this work aims to demonstrate the ET method as the pre-treatment for microalga lipid extraction and investigating the effect of Electrolysis Treatment Time (ETT), Electrolysis Field Strength (EFS) and Electrolysis Recycling Flowrate (ERF) on the lipid extraction of *Ankistrodesmus* sp. However, erstwhile to this investigation, the determination of the best ET systems in the combinations of recycle, without recycle, aeration and/or without aeration, was employed. The types of Fatty Acid Methyl Esters (FAME) in *Ankistrodesmus* sp. after the ET extraction was also determined, and the amount of methyl palmitate (C16:0) extracted was revealed in this study.

II. MATERIALS AND METHOD

A. Microalga Cultures

Ankistrodesmus sp. was obtained from Borneo Marine Research Institute (BMRI), Universiti Malaysia Sabah, Malaysia. The microalga was cultured phototrophically in sterilized freshwater enriched with Bold's Basal medium (BBM). The cells length were between 14.7-38.1µm (Mansa *et al.*, 2018a). The microalga was pre-cultured in a 500ml, 2000ml and 5000ml Erlenmeyer flask before it was up-scaled into a 10,000ml vessel at $32.1 \pm 0.7^\circ\text{C}$, pH 8.02 ± 0.07 and 12:12 h light-dark cycle under a light intensity of 999 ± 15 Lux for seven days of cultivation.

B. Extraction Procedures

1. Electrolysis treatment chamber design

The schematic diagram of ET chamber was shown in Figure 1. The electrolysis treatment (ET) chamber was fabricated

from an acrylic material with dimensions of 0.179m (length) x 0.152m (height) x 0.057m (width) and 0.005m (thickness). The stainless steel plate electrode dimensions were 0.153m (length) x 0.114m (height) and 0.0004m (thickness). Four bolts with five hexagonal nuts and four flat washers were used as the holder for the stainless steel plate electrode. The gap between the electrodes was fixed at 0.015m. Straight pipe fitting tubes were used for the inlet/outlet connector of the ET chamber. Four pneumatic flow control valves, hose to hose connector and 2.2m silicone tube were used so that the microalga can be recycled back to the ET chamber. The inlet and outlet flow were placed at the same level.

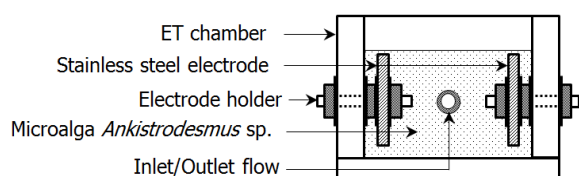


Figure 1. Schematic diagram of the ET chamber from the front view

2. Electrolysis treatment experimental setup

i. Selection of ET system

Three systems of ET were varied based on the combinations of aeration, without aeration, recycle and/or without recycle. The stainless steel plates were connected to a DC power supply (MP 303-3, Meguro). For the recycling and aeration flow, a peristaltic pump (Pump Drive PD 5006, Heidolph) and an air pump (AC-9980, Resun®) were used, respectively. Meanwhile, the sample flow rate was measured using a digital flow rate meter (T14 Flo-meter, McMillan Co.). Four types of treatment conditions for the ET system were carried out; (1) condition was without ET, Recycling Flowrate (RF) and Aeration Flow (AF); (2) condition (ET system 1) which undergone the ET only; (3) condition (ET system 2) which undergone the ET with RF only; (4) condition (ET system 3) which undergone the ET with RF and AF. The ETT, EFS, RF, AF, microalga volume and concentration were fixed at 25 min, 21V/cm, 140ml/min, 100ml/min, 1000ml and ~1.00g/l, respectively. All treatments were conducted in triplicates, and the data presented were in average value \pm standard errors.

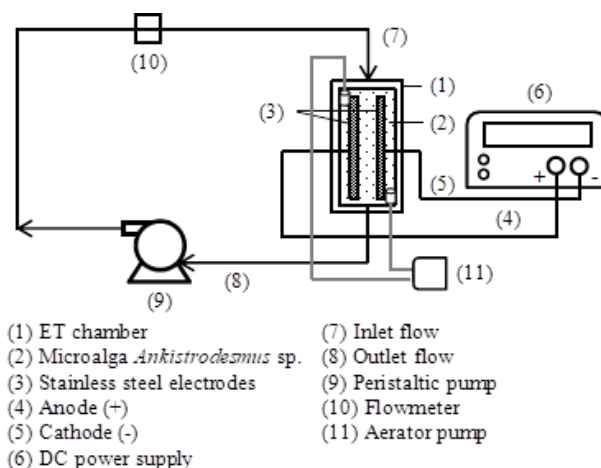


Figure 2. Schematic diagram of the experimental setup for the ET method

ii. Experimental design parameters

The schematic diagram of ET set up, as shown in Figure 2. Then, solvent extraction was carried out for further lipid extraction. The first parameter conducted was the ETT and varied at 10, 15, 20, 25, 30, 35 and 40 min at constant EFS (21V/cm) and ERF (140ml/min). The second parameter was the EFS and varied at 6, 9, 12, 15, 18 and 21V/cm at constant ETT (20 min) and ERF (140ml/min). The third parameter was the ERF and varied at 140, 160, 180, 200, 220 and 240ml/min at constant ETT (20 min) and EFS (21V/cm). The highest lipid obtained in ETT condition was selected and used in EFS. The same procedure was applied in the ERF experiment. Solvent extraction was carried out after the ETT, EFS and ERF experiment for further lipid extraction. All the above experiments were performed in triplicates, and the data presented were average value \pm standard errors.

3. Solvent extraction

Ankistrodesmus sp. lipid was extracted from its dried biomass according to Folch method (Folch *et al.*, 1957). After the ET method, all samples were centrifuged (3-18K Sartorius, Sigma) at 8,000rpm for 15 min and dried into an oven (UNE 200, Memmert) at $70 \pm 1^\circ\text{C}$ for 24 h. The dried biomass was placed inside a desiccator and then left for a few minutes for cooling at room temperature. A solvent mixture of 8ml chloroform and 4ml methanol ($\text{CHCl}_3\text{-MeOH}$; 2:1; v/v) were mixed with the dried microalga using a magnetic stirrer (HS0707V2, Favorit) at 850rpm for 10

min. Then, the biomass (solid phase) was filtered using a filter paper. Meanwhile, the liquid phase (a mixture of CHCl_3 -MeOH) was placed into a new clean universal bottle and 2.4ml distilled water was added into the liquid phase for washing. The liquid phase contained water was stirred at 1150rpm for 1 min to allow the formation of the lipid-solvent molecules and at 850rpm for 4 min for the two layers formation. The bottom layer consisted of CHCl_3 and lipid, whereas the top layer consisted of MeOH-water and non-lipids. The bottom layer was collected using a glass Pasteur pipette and placed into a new clean universal bottle.

4. The potential difference of cell membrane estimation

The potential difference of the cell membrane can be estimated using Equation 1 (Ortega-Rivas, 2012).

$$Um = \alpha d_c E \cos \theta \quad (1)$$

Where;

The potential difference of the cell membrane = U_m (V)

The parameter based on cell shape = α (dimensionless)

The cell diameter = d_c (μm)

The external electric field = E (V/cm)

The angle between the sites on the cell membrane = θ

5. The potential difference of cell membrane estimation

i. Light microscopic

A light microscope (Eclipse 90i, Nikon) was used to view the *Ankistrodesmus* sp. cells before and after the ET method. An approximately 10 ml of each sample was taken for cells observation. The magnification power used to view the microalga cells was 40X.

ii. Scanning electron microscopy (SEM)

The surface morphology of the *Ankistrodesmus* sp. cells were examined by Scanning Electron Microscopy (SEM) (Carl Zeiss, EVO MA 10). The SEM sample preparation was performed based on chemical fixation method, and the procedures were followed according to Mansa *et al.* (2018a).

6. Lipid content determination

The lipid content was determined based on the gravimetric method. The procedures were followed according to Mansa

et al. (2018a). The lipid content was calculated using Equation 2.

$$\text{Lipid content (\% wt)} = \left[\frac{\text{glipid microalga}}{\text{gdried microalga}} \right] \times 100\% \quad (2)$$

7. Lipid functional group determination

A Fourier Transform Infrared Spectroscopy (FTIR) (2000 Series, Perkin-Elmer) was used to determine the functional groups of lipid for all samples. The procedures were followed according to Mansa *et al.* (2018a).

8. Transesterification process

A base catalyst (potassium hydroxide, KOH) was used for the FAME conversion, and the procedures were followed according to Mansa *et al.* (2018a).

9. Fatty acid methyl ester composition

A Gas Chromatography-Mass Spectrophotometer (GCMS) (Model 6890N, Agilent Technologies) was used to determine the FAME compositions. The procedures were followed according to Mansa *et al.* (2018a).

III. RESULTS AND DISCUSSION

A. Lipid Functional Groups of *Ankistrodesmus* sp.

1. In ET system varied conditions

The FTIR spectrum for each sample was shown in Figure 3. The lipid functional groups were all detected in each sample for solvent extraction without ET and with ET method (ET system 1, 2 and 3). The C-H stretching vibration was obtained at 3010cm^{-1} representing the stretching of $=\text{C}-\text{H}$ bond, at 2855cm^{-1} , 2925cm^{-1} and 2954cm^{-1} representing the stretching of C-H bonds in $-\text{CH}_3$ and CH_2 . It was presented that, C=O bond found in the ester group was detected at 1740cm^{-1} , which was primarily obtained from lipid and fatty acids compound (Dean *et al.*, 2010). Besides that, C-O bond and C-O-C bond was also detected at 1380cm^{-1} and $1250-1070\text{cm}^{-1}$ which also came from the ester group, respectively (Forfang *et al.*, 2017). The peak at 1460cm^{-1} was also

obtained, indicating the bending of CH₂ bond (Bartošová *et al.*, 2015).

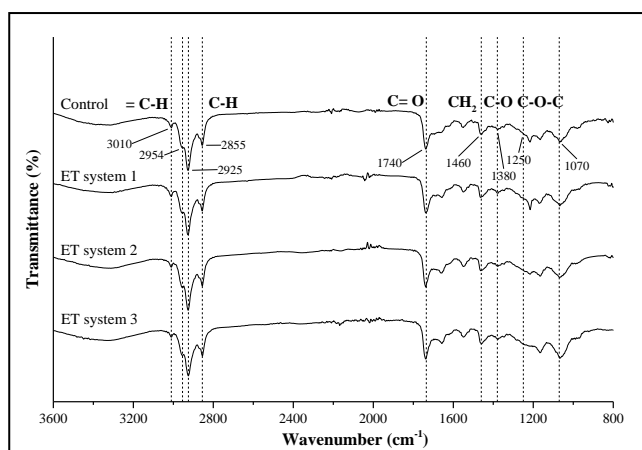


Figure 3. FTIR spectra of *Ankistrodesmus* sp. in the control sample and different ET system

2. In ETT, EFS and ERF condition

The FTIR spectra for ETT, EFS and ERF performed solvent extraction with the ET method were displayed in Figure 4. Figure 4a, 4b and 4c showed the FTIR spectra from ETT, EFS and ERF samples, respectively. The lipid functional groups were found to be similar to the lipid profiles presented in Figure 3. In Figure 4a, the highest peak intensity was obtained from ETT20 followed by ETT25, ETT30, ETT35, ETT40, ETT15, ETT10 and ETT0. Meanwhile, the highest peak intensity was obtained from EFS21 followed by EFS18, EFS15, EFS12, EFS9, EFS6 and ETT0, as shown in Figure 4b. For EFS, the highest peak intensity was obtained from ERF180 followed by ERF200, ERF220, ERF240, ERF160, ERF140 and ERF0. From these FTIR spectra, it shows that higher peak intensity corresponded to the higher lipid content that was extracted from the sample (Forfang *et al.*, 2017).

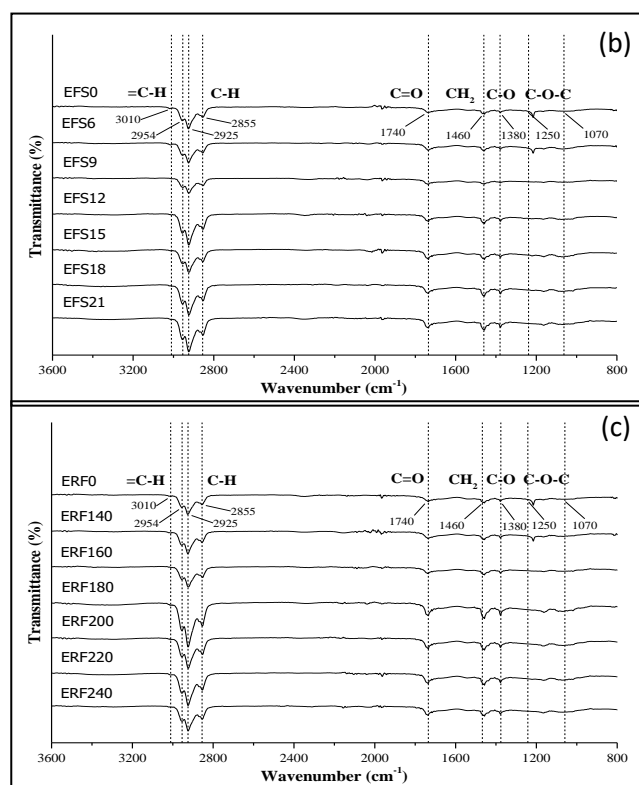
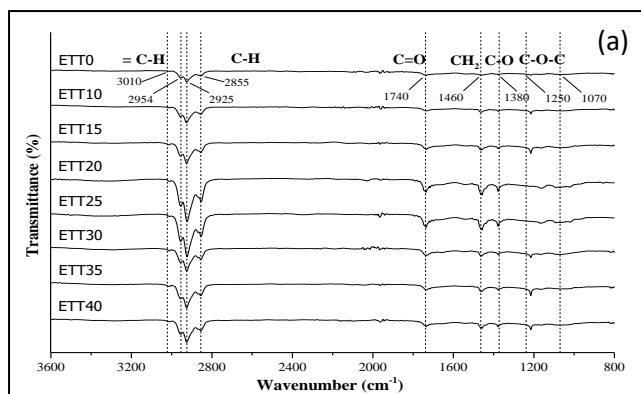


Figure 4. FTIR spectra of *Ankistrodesmus* sp. in (a) ETT, (b) EFS and (c) ERF samples

B. Lipid Content

1. Effect of ET

The effect of the ET system on the lipid content of *Ankistrodesmus* sp. was shown in Figure 5a. Based on Figure 5a, the highest lipid content extracted was obtained from ET system 3 with $13.05 \pm 0.57\%$ wt. followed by ET system 2, the control sample, and ET system 1 with $12.91 \pm 0.85\%$ wt., $11.15 \pm 0.73\%$ wt. and $10.26 \pm 0.41\%$ wt., respectively. Except for ET system 1, all samples that had undergone the ET method showed higher lipid content compared to the control sample.

Higher lipid content was attained from the ET system three due to RF and AF systems were introduced into the system. The microalga cells were well mixed and had higher chances of passing through the electric field region. Thus, it provided a higher probability of the cell membrane to be disturbed or weakened. This facilitated the solvent molecules to penetrate deeper into the cell and allow it to extract the lipid that stored inside the lipid granule (Eing *et al.*, 2013; Guldhe *et al.*, 2016). In ET system 2, only RF was introduced, thereby, explained the difference in lipid

content extracted from ET system 3. However, the lipid content obtained from the ET system 1 was lower compared to the control sample. The reasonable explanation for this result was because of the oxidation of lipids may be occurred along with the experiment (Daghrir *et al.*, 2014). Apart from the surrounding oxygen gas, the ET also produced oxygen gases at the anode. Thus it can contribute to the increase of lipid oxidation. The lipid oxidation can also occur in ET system 2 and system three as the anode electrode undergoes an oxidation process while the cathode undergoes the reduction process.

However, the condition in the ET system one might be an extreme condition for the *Ankistrodesmus* sp. lipid

extraction as compared to the other two methods. Misra *et al.* (2014) had discovered that using an electrochemical treatment (non-sacrificial carbon electrode) the final lipid content of *Chlorella sorokiniana* (12.20% wt.) and *Scenedesmus obliquus* (15.42% wt.) were 0.17% and 0.14% lower than the control sample, respectively. This indicates that using ET without AF and RF do not improve the lipid extraction but can contribute to lower lipid extraction with only slightly different. Mansa *et al.* (2018b) had observed that using ET with AF, the final lipid attained of *Chlorella* sp. was 0.86% or 1.11 times lower than the WOET sample. The effect ET system towards the cell appearances and amount of extracted lipid were summarized in Table 1.

Table 1. Cell observation and total lipid based on different ET system

System	Control	System 1	System 2	System 3
Treatment system conditions	without ET, RF and AF	with ET but without RF and AF	with ET and RF but without AF	with ET, RF and AF
Batch/Semi-continuous system	-	batch	semi-continuous	semi-continuous
Final temperature (°C)	25.1 ± 0.1	48.4 ± 0.1	45.4 ± 0.1	43.0 ± 0.1
Final pH	8.55 ± 0.01	10.12 ± 0.01	9.21 ± 0.01	8.94 ± 0.01
Conductivity (μS/cm)	1160 ± 1	1966 ± 3	1809 ± 2	1792 ± 3
The cells appearance after the ET method	Non-flocculate and non-fluffy-like	flocculate and fluffy-like	fluffy-like	fluffy-like
Lipid content (% wt.)	11.15 ± 0.73	10.26 ± 0.41	12.91 ± 0.85	13.05 ± 0.57

2. Effect of ETT

The effect of ETT on the extraction of microalga lipid from zero to 40 min was displayed in Figure 5b. The lipid content obtained from sample WOET (ETT0) was 11.83 ± 0.81% wt. All samples WET (ETT10 – ETT40) had produced higher lipid content than sample WOET. According to these chart bars, the lipid content extracted from *Ankistrodesmus* sp. increased from 10 to 20 min. However, the lipid content had decreased from 20 to 40 min. The amount of lipid content extracted was the highest at 20 min, with 15.83 ± 0.68% wt. and lowest at 10 min with 13.59 ± 0.94% wt.

The entire lipid content attained in samples WET were higher than sample WOET due to ET was used as pre-

extraction. Increasing the PEF treatment time will also rapidly increase the rate of microbes' inactivation, then slowly plateau and further treatment will show no significant change (Huang & Wang, 2009). Hence, to correlate the above statement with the present study, the extraction of *Ankistrodesmus* sp. lipid increased from 10 to 20 min because, during this period, the cell's membrane was experienced vigorous irreversible electroporation. The lowest lipid content attained was at 10 min because the cells had the lowest exposure time. Meanwhile, the highest lipid content extracted was at 20 min and extraction duration longer than 20 min will cause an extreme extraction process, which contributes to the decrease of lipid content extraction from 20 to 40 min. The results obtained here agreed with

other reported literatures. As reported by Daghrir *et al.* (2014), increasing the electrochemical treatment time between 60 to 100 min will enhance *C. vulgaris* lipid content between 3.33 to 3.55% wt., respectively. In fact, by increasing the PEF treatment time between 1 to 3 min had shown to increase the percentage of *Chlorella* sp. disrupted cells from 40 to 90% (Rego *et al.*, 2015).

Unfortunately, increasing the treatment time above than 20 min, the lipid content decreased. The possible explanation for this condition was because of the lipid being oxidized. In theory, at a certain point, further treatment of the microbe inactivation will show no significant change (Huang & Wang, 2009). However, because of the oxidation mechanism due to the presence of oxygen gases either from the surrounding or released from the anode terminal, will cause some of the extracted lipid to oxidize. The degradation of lipid can be due to hydrolysis or oxidation mechanism, and this can be attributed by photo-oxidation, auto-oxidation or enzymatic-oxidation reactions (Meullemiestre *et al.*, 2016). It is also essential to take note that, when the treatment time is increased, the current intensity also increase and this had will cause the sample's temperature to rise. Thereby, it may affect the valuable components such as lipid that released into the surrounding medium to oxidize (Daghrir *et al.*, 2014; Mansa *et al.*, 2018b). This may indicate that the lipid has undergone an oxidation process. Therefore, this lipid oxidation may attribute to the lowering of *Ankistrodesmus* sp. lipid content.

3. Effect of EFS

The effect of EFS on the extraction of microalga lipid is shown in Figure 5c. The lipid content gained in sample WOET (EFS₀) was $11.19 \pm 0.76\%$ wt. As expected, if the EFS value increased the lipid content of *Ankistrodesmus* sp. extracted was also increased. Same as in ETT, the lipid content from the samples WET (EFS₆ – EFS₂₁) were higher than the sample WOET. The highest lipid content was achieved at 21V/cm with $15.46 \pm 0.78\%$ wt. and the lowest lipid content was at 6V/cm with $11.36 \pm 0.79\%$ wt.

As stated by Donsì and co-workers, most of the cell membrane has a critical transmembrane potential around 0.2 to 1.0V (Donsì *et al.*, 2010). However, the cell

membrane potential calculated in this study was much lower than 0.2V. The potential difference of the cell membrane of microalga (U_m) can be estimated using the Equation 1 ($U_m = \alpha d E_{cos \Theta}$). In this study, the structure of *Ankistrodesmus* sp. cells were observed to have a needle-like shape and curvy or sharp at both edges as reported by Mansa *et al.* (2018b). By considering the *Ankistrodesmus* sp. cell with almost a rectangular shape ($\alpha=1.0$; considering the middle part of the cell) and the cell width (0.3-2 μ m; 0.3 μ m is based on literature, and 2 μ m is the width obtained in this study see Figure 7a), the potential difference can be attained around 1.8×10^{-4} to 1.2×10^{-3} V. Thus, using 6V/cm and above can disrupt most of the cells.

Besides that, the result attained here was supported by other reported literatures. Heinz *et al.* (2003) stated that increasing the electric field strength will also increase treatment efficiency. Applying higher field strength on the microalga (above the critical strength of the cell membrane) will increase the cell membrane permeability by making the cell membrane to become irreversible electroporation (Joannes *et al.*, 2015b; Zbinden *et al.*, 2013; Zimmermann *et al.*, 1974). This will assist the solvent molecules that were added to penetrate the microalga's cell. Hence, this explained that the lipid content extracted has increased as the EFS increased. Based on the graph, the optimum value for EFS has not yet achieved, because the highest voltage that can be used was up to 31.4V only, due to the equipment limitation (i.e. DC power supply). Higher EFS (≥ 21 V/cm) can be obtained if the gap between the two electrodes (d) is reduced. However, in this study, the smallest gap used was 0.015m and was fixed for the entire experiment.

4. Effect of ERF

The effect of ERF on the extraction of microalga lipid was shown in Figure 5d. The lipid content attained in sample WOET (ERF₀) was $11.21 \pm 0.83\%$ wt. Similar to the ETT and ERF, all the samples WET (ERF₁₄₀-ERF₂₄₀) had produced higher lipid content than in sample WOET. According to the chart bars, the lipid content extracted from *Ankistrodesmus* sp. increased from 140 to 180ml/min. Nevertheless, the lipid content had decreased from 180 to 240ml/min. The amount of lipid content extracted was the highest at 180ml/min of

recycling flowrate with $15.60 \pm 0.67\%$ wt. and lowest at 140ml/min of recycling flowrate with $12.32 \pm 0.94\%$ wt.

The entire lipid content gained from samples WET (ERF140-ERF240) were also found to be higher than the sample WOET. From these chart bars, increasing the ERF at a certain level will also increase the lipid extraction efficiency. The results gained in this study agreed with Daghrir and co-workers. They had investigated the effect of recycling flowrate towards the *Chlorella vulgaris* lipid extraction. At 340ml/min, the lipid obtained was 3.33% wt., whereas at 440ml/min lipid obtained was increased to 4.81% wt. at constant 0.5A for 60 min (Daghrir *et al.*, 2014). Introducing the recycling flow in the ET has given advantages of the microalga cells to pass through the electric field region and enhanced the mass transfer coefficient. Furthermore, by increasing the recycling flow is capable of reducing the diffusion layer thickness near the anode (decrease the microalga cells precipitation), therefore provides better performance of the ET (Straessner *et al.*, 2016).

Even so, increasing the ERF above than 180ml/min had led to the decreasing of extraction of the microalga lipid content. Some cell disruption methods such as bead mill had shown a reduction of cell disruption of *Saccharomyces cerevisiae* if high flowrate was employed (Middelberg, 1995). This was because, when the feed flow rate increases the residence time of the microalga cells to pass through the electric field region will be reduced. Thereby, it may decrease the contact time of *Ankistrodesmus* sp. cells to be exposed by these electric fields. By means, high possibility of reversible electroporation mechanism will occur. In this way, ERF may affect the lipid extraction process.

Note that, the amount of lipid content obtained from the control samples (ETTo, EFSO and ERFO) were all different because all the microalga cells were harvested at the same day, but the lipid extraction was not performed at the same day. The ET method was carried out one at a time and not in a parallel experiment. For each parameter, including the replicates (from ET method to FAME determination experiment), it required at least five days to be completed. The first parameter conducted was the ETT, followed by EFS and ERF. Because of this, it was suspected that the lipid content of microalga might be changed due to this

condition. Therefore, for each parameter, it has its control sample, as this was to ensure the culture sample for each parameter must be the same condition as possible and avoiding from attaining bias results.

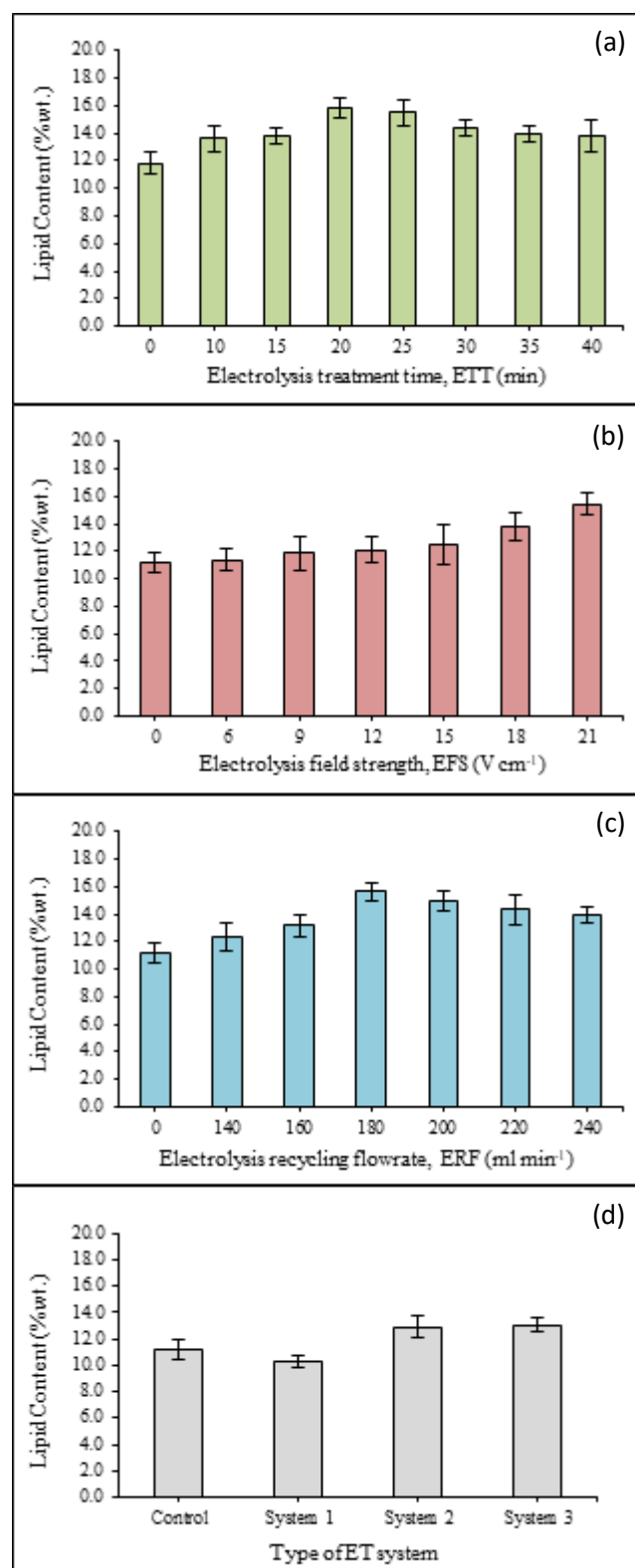


Figure 5. Effect of (a) ET systems, (b) ETT, (c) EFS and (d) ERF on the lipid extraction of *Ankistrodesmus* sp.

C. Comparison of Other Alternative Extraction Processes

The comparison of other alternative lipid extraction processes with several of microalgae species are presented in Table 2. This includes the ET method, electrochemical treatment, PEF, microwave irradiation, ultrasonication, hydrothermal treatment and Soxhlet extraction. From Table 2, it was found that of all these methods the hydrothermal treatment has demonstrated to provide the highest lipid extraction with 84% wt. of *Nannochloropsis* sp. that was approximately 54% of lipid increment (Qu *et al.*, 2018). This can be attributed by using high temperature (i.e. 180°C) which can certainly destroy the cell wall of microalga. However, using this method may change the colour of microalga from dark green to brown and longer treatment time is needed (i.e. 60 min) to achieve the highest lipid extraction. Other researchers like Yao *et al.* (2018) had also used an ultrasonication method to extract *Nannochloropsis* sp. lipid. It was reported that the extracted lipid was $73.2 \pm 8.4\%$ wt. This method is currently under developing for large scale production.

Other than this extraction method, it was found that using the reactive electrochemical had improved the lipid extraction of *S. dimorphus* up to $23.4 \pm 0.7\%$ wt. with 8.2% of lipid increased (Hua *et al.*, 2016). The type of electrode material used (i.e. Ti_4O_7) is one of the factors that affect the lipid extraction performance. On the other hand, to convert Ti_4O_7 from TiO_2 involves very high temperature at 900°C, which extremely dangerous and requires a proper place for experimenting. According to Bensalem *et al.* (2018), the lipid of *C. reinhardtii* can be extracted up to $88.2 \pm 2.0\%$, which an increased about 6.8% as compared to the PEF untreated sample. The findings also revealed that the cell

disruption method, such as PEF is necessary prior to solvent extraction for extracting the lipid from *C. reinhardtii*. Nevertheless, it was also discovered that exposing 5.5Kv/cm on the cell did not lyse or weaken the cell wall of *C. reinhardtii* (wild type strain).

Meanwhile, *Tetraselmis* sp. lipid can be extracted around 5.4% at 65°C for 10 min using the microwave irradiation assisted extraction (Teo and Idris, 2014). The small lipid content might due to the small volume of the sample was used in their study. However, by comparing with the present study, the lipid extracted percentage here was 2.9 times higher. Interestingly, some of the extraction methods presented in Table 2 such as Soxhlet extraction and electrochemical treatment have shown lower lipid extraction than the untreated microalgae (Daghrir *et al.*, 2014; Escorsim *et al.*, 2018). *A. obliquus* and *C. vulgaris* lipid can be extracted at $6.76 \pm 0.17\%$ wt. and $5.53 \pm 0.23\%$ wt. using Soxhlet extraction and electrochemical treatment, respectively. However, the final lipid extracted were both slightly lower than the untreated sample. The previous study conducted by Mansa *et al.* (2018b) also reported that the lipid extracted from *Chlorella* sp. at 14V/cm for 50 min using electrolysis treatment was slightly lower than the ET-untreated microalga.

Based on Table 2, it is shown that using different lipid extraction method with a variety of microalga species will offer dissimilar extraction performances. However, the findings obtained here are comparable with other extraction methods (i.e. Soxhlet extraction, microwave irradiation and electrochemical treatment). Perhaps, further investigation regards to the volume of microalga need be done for this ET method to become practicable in a large scale application for biodiesel production.

Table 2. Comparison of the proposed ET method with other alternative extraction processes

Methods	Parameters	Optimum values	Reference
Electrolysis Treatment (ET)	Microalga species	<i>Ankistrodesmus</i> sp.	Present study
	Initial lipid content	$11.21 \pm 0.83\%$ wt.	
	Final lipid content	$15.60 \pm 0.67\%$ wt.	
	Field strength	21V/cm	
	Current intensity	$2.50 \pm 0.01A$	
	Treatment time	20 min	
	Recycling flow	180ml/min	

	Microalga conc.	1.00 ± 0.03g/l	
	Electrode type	Stainless steel (cathode and anode)	
	Solvent extraction	Folch method	
Electrolysis Treatment (ET)	Microalga species	<i>Chlorella</i> sp.	(Mansa <i>et al.</i> , 2018b)
	Initial lipid content	8.58 ± 0.49% wt.	
	Final lipid content	7.72 ± 0.54% wt.	
	Field strength	14V/cm	
	Current intensity	1.92 ± 0.02A	
	Treatment time	50 min	
	Recycling flow	140ml/min	
	Microalga conc.	0.46 ± 0.01g/l	
	Electrode type	Stainless steel (cathode and anode)	
	Solvent extraction	Folch method	
Reactive Electrochemical	Microalga species	<i>Scenedesmus dimorphus</i>	(Hua <i>et al.</i> , 2016)
	Initial lipid content	15.2 ± 0.6% wt.	
	Final lipid content	23.4 ± 0.7% wt.	
	Field strength	8V/cm	
	Current intensity	0.5A	
	Treatment time	120 min	
	Microalga conc.	1.4 g/l	
	Electrode type	Stainless steel (cathode) and Ti ₄ O ₇ (anode)	
	Solvent extraction	Dichloromethane and methanol	
Electrochemical	Microalga species	<i>Chlorella vulgaris</i>	(Daghrir <i>et al.</i> , 2014)
	Initial lipid content	6.23 ± 1.05% wt.	
	Final lipid content	5.53 ± 0.23% wt.	
	Field strength	14.3V/cm	
	Current intensity	0.6A	
	Treatment time	100 min	
	Recycling flow	394.51ml/min	
	Electrode type	Stainless steel (cathode) and Ti/IrO ₂ (anode)	
	Solvent extraction	Folch method	
	Electrolyte	1.4g/l of Na ₂ SO ₄	
Pulsed Electric Field (PEF)	Microalga species	<i>Chlamydomonas reinhardtii</i>	(Bensalem <i>et al.</i> , 2018)
	Initial lipid content	81.4 ± 2.3% ^a	
	Final lipid content	88.2 ± 2.0% ^a	
	Field strength	5.5Kv/cm	
	Frequency	10Hz	
	Pulse duration	5μs	
	Number of pulses	10 unipolar	
	Microalga volume	130μl (3×10 ⁶ cells/ml)	
	Electrode type	Stainless steel (cathode and anode)	
	Solvent extraction	Hexane	

Microwave Irradiation	Microalga species	<i>Tetraselmis</i> sp.	(Teo and Idris, 2014)
	Initial lipid content	~5.0% ^b	
	Final lipid content	~5.4% ^b	
	Power	500W	
	Treatment	65°C	
	Temperature	10 min	
	Treatment time	15ml	
	Microalga volume	Folch Method	
	Solvent extraction		
Ultrasonication	Microalga species	<i>Nannochloropsis</i> sp.	(Yao <i>et al.</i> , 2018)
	Initial lipid content	Not stated	
	Final lipid content	73.2 ± 8.4% wt. ^c	
	Frequency	20kHz	
	Power density	3.8W/ml	
	Treatment time	5 min	
	Microalga mass	3g (12% solid conc.)	
	Solvent extraction	n-hexane	
Hydrothermal Treatment	Microalga species	<i>Nannochloropsis</i> sp.	(Qu <i>et al.</i> , 2018)
	Initial lipid content	30.44 ± 1.46% wt.	
	Final lipid content	84% wt.	
	Treatment temperature	180°C	
	Treatment time	60 min	
	Microalga conc.	20% wt. solid conc.	
	Solvent extraction	Diethyl ether	
Soxhlet Extraction	Microalga species	<i>Acutodesmus obliquus</i>	(Escorsim <i>et al.</i> , 2018)
	Initial lipid content	7.41 ± 0.16% wt.	
	Final lipid content	6.76 ± 0.17% wt.	
	Treatment time	12 h	
	Microalga mass	20g	
	Solvent	Ethanol: Hexane (1:2; v/v)	

Note: ^a lipid extraction is expressed based on absorbance; ^b lipid extraction is expressed in volume extracted lipid (ml) / cell dry weight (g); ^c quantified based on the non-polar lipid

D. Microscopy Results

1. Microscopic cell observation

The *Ankistrodesmus* sp. cells samples before and after the ET method was viewed under a light microscope. This was to observe any physical changes, such as shapes or abnormal appearances. The microscopic images reported in this study were based on the highest lipid content attained in each ET parameter from ETT20, EFS21 and ERF180 sample. The microscopic images for *Ankistrodesmus* sp.

cells before the ET method were shown in Figure 6a. As observed, the *Ankistrodesmus* sp. cells have needle-like shaped in green colour and were found to be similar to other reported literature (Mansa *et al.*, 2018a; Sukkrom, 2014).

Figure 6b, Figure 6c and Figure 6d displayed the microscopic images for *Ankistrodesmus* sp. cells after the ET method and were taken from the ETT20, EFS21 and ERF180 sample, respectively. Based on these microscopic images, the cells prior to ET method were intact (see Figure 6a), and after the ET method, some cells did not have the

needle-like form. Moreover, flocculated green materials without cell membranes were observed. These green materials were believed to be the intracellular material or the green pigment of *Ankistrodesmus* sp. Noted that, the sample's conductivity increased during the ET experiment

(data not shown). The increasing conductivity indicates the presence of ionic compounds such as lipids, carbohydrates and proteins (Postma *et al.*, 2016). Therefore, this shows that *Ankistrodesmus* sp. cell membrane may have been disrupted during the ET method.

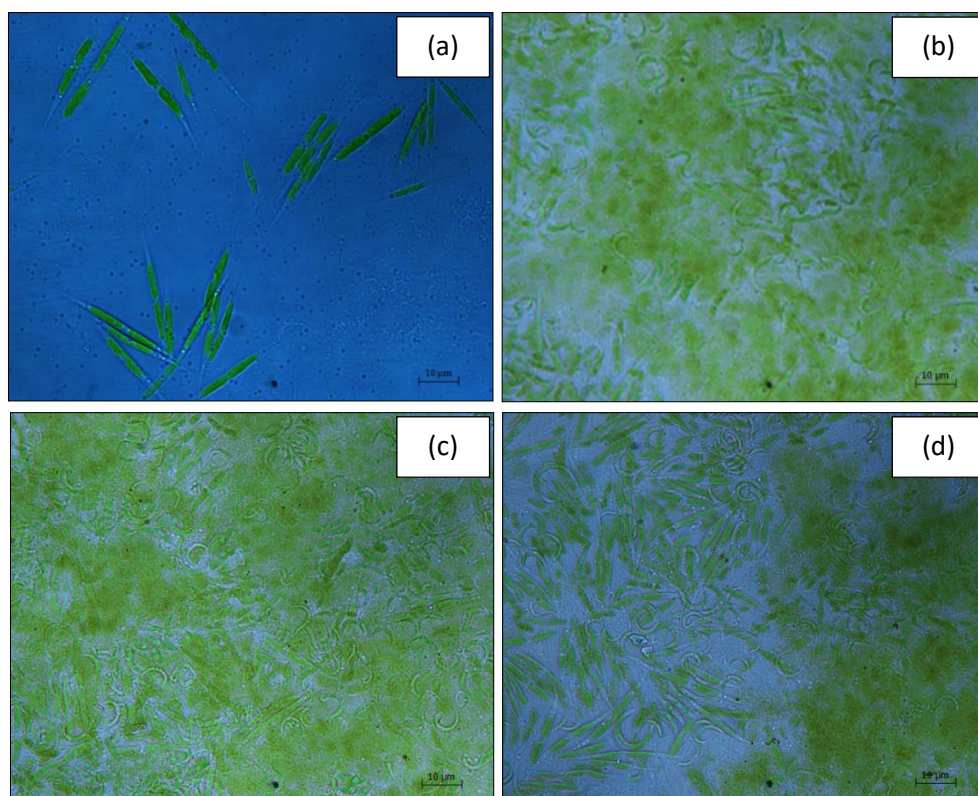


Figure 6. *Ankistrodesmus* sp. cells are taken from (a) sample WOET, (b), ETT20 (c) EFS21 and (d) ERF180 samples viewed under a light microscope at 40X magnification power

2. Microalga cells morphology

To support the microscopic observation results, the condition of *Ankistrodesmus* sp. cells before and after the ET method were also viewed under SEM to observe if any cells changes in term of sizes, shapes or appearances. The SEM images were taken from ETT20, EFS21 and ERF180 sample (based on the higher lipid content extraction). Figure 7a showed the SEM images for sample WOET and was viewed at 3000X magnification power. The cells have a needle-like shape and curvy or sharp extremes, as presented in Figure 6a. Moreover, some of the cells were observed to have irregular crests (see the white arrows) as shown in Figure 7b. The cell morphology observed here were similar to *Ankistrodesmus gracilis* (Asselborn *et al.*, 2015).

Figure 7b shows the SEM images from the ETT20 sample.

Based on these SEM images, some cells do not have their cell membrane (white arrows). The cells without membrane might be the green materials without membrane that were observed from the microscopic observation. These fragmented and agglomerated materials were believed to be the intracellular components of *Ankistrodesmus* sp., which were similar to *Chlorella* sp. (Yap *et al.*, 2014). This shows that *Ankistrodesmus* sp. cells had been disrupted by the ET method.

Figure 7c shows the SEM images from the EFS21 sample. The images taken were also similar to Figure 7b, which some cells do not have their cell membrane (white arrows). It is believed to be the green materials without its cell membrane. Figure 7d shows the SEM images from the ERF0.18 sample. Based on these SEM images, it was observed that some cells were fragmented and adhered with each other (see the white

arrows). In addition, some cells have no sharp edges (see the white arrows), which were likely broken. Hence, these

SEM images supported the results obtained from the microscopic observation.

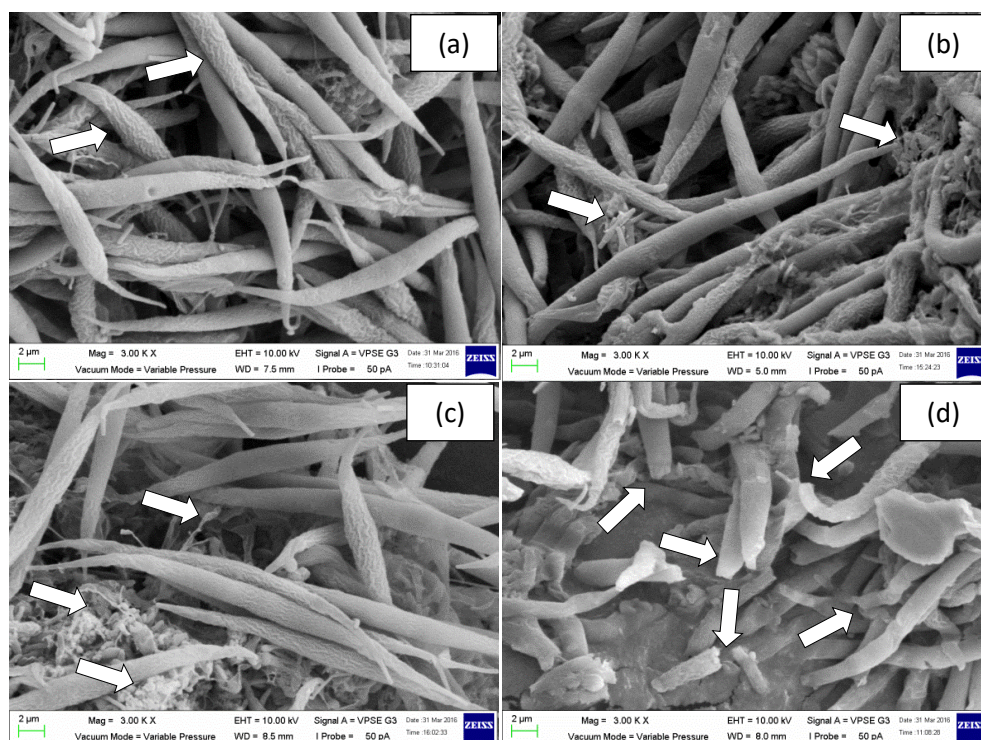


Figure 7. *Ankistrodesmus* sp. cells morphology taken from (a) sample WOET, (b) ETT20, (c) EFS21 and (d) ERF180 samples viewed under SEM at 3000X magnification power

D. Fatty Acid Methyl Ester Compositions

There were six significant peaks obtained from the GCMS chromatogram at the retention time of 16.08, 16.24, 16.95, 22.18, 22.40 and 22.82. The compounds detected from these six peaks were summarised in Table 2. However, only four peaks were confirmed as the FAME compounds. The four FAME detected were 7,10-hexadecadienoic acid methyl ester, hexadecanoic acid methyl ester (methyl palmitate), (E,E)-9,12-Octadecadienoic acid methyl ester (methyl linoleate) and (Z,Z,Z)-9,12,15-octadecatrienoic acid,

methyl ester (methyl linolenate) at retention time 16.08, 16.95, 22.18 and 22.41, respectively. The number of peaks obtained here was the same as reported by Mansa *et al.* (2018a). From Table 3, among these six peaks, the compound detected at the retention time of 22.82 was dominated with 47.18%, which was the 3,7,11,15-tetramethyl-2-hexadecen-1-ol. Nevertheless, among the FAME compounds, methyl palmitate was the predominant compound with 15.78%, followed by methyl linolenate, 7,10-hexadecadienoic acid methyl ester and methyl linoleate with 14.28%, 6.84% and 6.51%, respectively.

Table 3. Compounds detected from *Ankistrodesmus* sp. WOET sample

No.	Compound Name	Retention Time (min)	Peak Area (%)
1.	7,10-hexadecadienoic acid methyl ester	16.08	6.84
2.	(Z,Z,Z)-9,12,15-Octadecatrien-1-ol	16.24	9.41
3.	Hexadecanoic acid methyl ester	16.95	15.78
4.	(E,E)-9,12-Octadecadienoic acid methyl ester	22.18	6.51
5.	(Z,Z,Z)-9,12,15-Octadecatrienoic acid methyl ester	22.41	14.28
6.	3,7,11,15-Tetramethyl-2-hexadecen-1-ol	22.82	47.18

Methyl palmitate consists of monosaturated fatty acids with carbon chain C16:0. Meanwhile, 7,10-hexadecadienoic acid methyl ester (two double bonds), methyl linoleate (two double bonds) and methyl linolenate (three double bonds) consist of polyunsaturated fatty acids with carbon chain C16:2, C18:2 and C18:3, respectively. The result obtained in this study agreed with the findings obtained by Sukkrom (2014), which has stated that palmitic acid (C16:0) was majorly found in *Ankistrodesmus* sp. lipid. The saturated fatty acids such as palmitic acid are favourable in biodiesel engine (Knothe & Razon, 2017). As for monounsaturated and polyunsaturated fatty acids, additional processes will be required to convert them into saturated fatty acids (Fuad *et al.*, 2015). Hence, adding an extra cost for the biodiesel conversion process.

The amount of methyl palmitate concentration in each ETT samples is shown in Figure 8a. For sample WOET, the methyl palmitate obtained was $0.113 \pm 0.018 \mu\text{g/ml}$. The methyl palmitate extracted was the highest at 20 min with $0.440 \pm 0.046 \mu\text{g/ml}$ and lowest at 10 min with $0.254 \pm 0.067 \mu\text{g/ml}$ at constant EFS (21V/cm) and ERF (140ml/min). The highest methyl palmitate attained in ETT20 can be correlated to the higher lipid content was also extracted at this point. Meanwhile, the decreasing of methyl palmitate can be attributed by lipid oxidation as previously discussed. Apart from that, several factors such as the type of fatty acids, oxygen, heat, photo-sensitivity and the existence of minor constituents (i.e. metals, phospholipids, free fatty acids, monoacylglycerol and triacylglycerol) can affect the chemical reactions in lipid to biodiesel (Meullemiestre *et al.*, 2016).

The amount of methyl palmitate concentration in each ETT samples was shown in Figure 8b. For sample WOET the methyl palmitate obtained was $0.145 \pm 0.010 \mu\text{g/ml}$. The methyl palmitate concentration extracted was the highest at 21V/cm with $0.629 \pm 0.058 \mu\text{g/ml}$ and lowest at 6V/cm with $0.250 \pm 0.040 \mu\text{g/ml}$ at constant ETT (20 min) and ERF (140ml/min). The highest methyl palmitate attained in EFS21 can be correlated to the higher lipid content was also extracted at this point. The amount of methyl palmitate concentration in each ERF samples was shown in Figure 8c. For sample WOET, the methyl palmitate obtained was $0.176 \pm 0.019 \mu\text{g/ml}$. The methyl palmitate extracted was

the highest at 180ml/min with $0.390 \pm 0.022 \mu\text{g/ml}$ and lowest at 140ml/min with $0.252 \pm 0.023 \mu\text{g/ml}$ at constant ETT (20 min) and EFS (21V/cm). The highest methyl palmitate attained in ERF21 can be correlated to the higher lipid content was also extracted at this point.

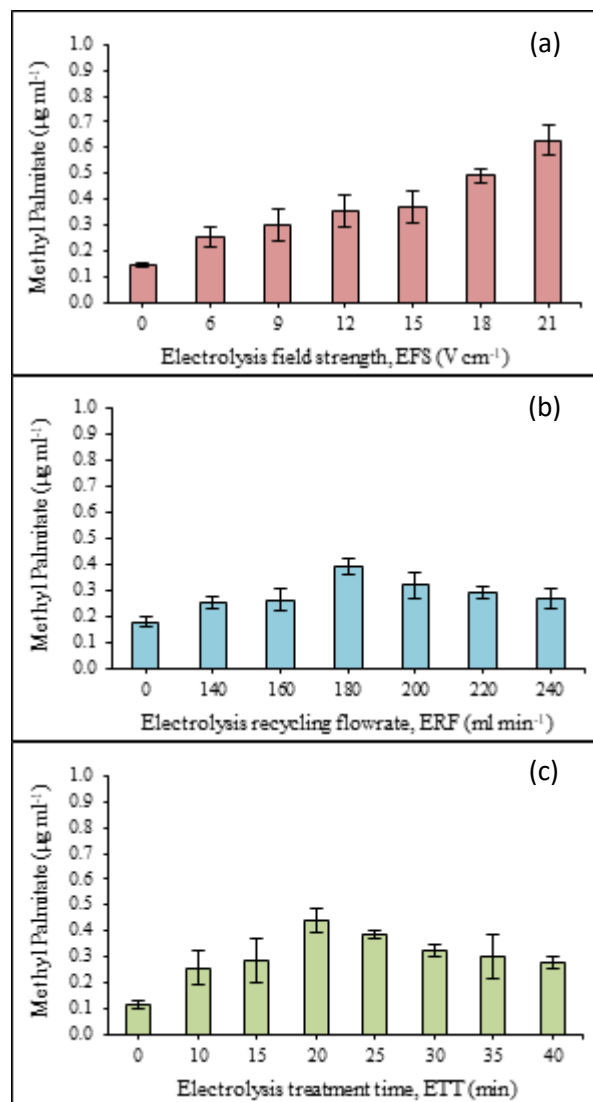


Figure 8. Effect of (a) ETT, (b) EFS and (c) ERF on the FAME of *Ankistrodesmus* sp.

IV. CONCLUSION

In this study, the effect of ETT, EFS and ERF towards the lipid extraction of *Ankistrodesmus* sp. were investigated. All samples from ETT, EFS and ERF that had undergone ET have produced higher lipid content than the control sample with an increase of lipid extraction from 1.34 to 1.40 times higher. Furthermore, it was shown that using the ET method prior to solvent extraction did not affect the FAME qualities

at all. Besides, it was also revealed that *Ankistrodesmus* sp. contained higher saturated fatty acids such as palmitic acid (C16:0), and this is a crucial feature in one of the biodiesel properties. This elucidated that using ET as the microalga pre-treatment for lipid extraction not only enhance the lipid extraction yield but also proven to sustain the quality of FAME compositions. Aside from being conventionally utilized in the microalga harvesting process, the ET could also be one of the alternative methods that can be used to distract the microalga's cell wall, especially for wet biomass in biodiesel production. The parameters such as the microalga concentration,

microalga species types, solvent types and electrode types can be investigated for the improvement of lipid extraction using the ET method.

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